

Treating Neurodegenerative Disorders: Towards Detailed Pathophysiology and Precision Medicine

Katie Gu

Neurodegenerative diseases (NDs) encompass a wide range of acute and chronic conditions characterised by the loss of neuron and glial cells in the brain or spinal cord. In acute conditions, due to stroke or spinal cord injury, different types of neurons within a restricted brain region die over a short amount of time. In chronic conditions, there is either a selective neuronal loss of specific sub-populations (such as dopamine neurons in PD or medium spiny neuron loss in HD), or widespread degradation of many neuronal types (such as in AD) over several years. For both types, no cures currently exist. Current clinical approaches adopted can only target symptomatic treatment and disease management, while clinical

applications of novel therapies developed for NDs have been limited.

In this paper, I review the current literature on both implemented and developing therapies for Alzheimer's (AD), Parkinson's (PD), and Huntington's (HD) disease, three of the most common and debilitating chronic NDs. The symptoms, affected brain regions, and some molecular changes have been identified for several NDs, some of which are summarized in Table 1 . Novel therapies have been developed in animal models to target these observed pathological and molecular changes, but significant ethical, economic, regulatory, safety, and etiological weaknesses still hinder their widespread clinical application.

Table 1: Overview of Neurodegenerative Disorders

Disease	~Incidence (per year in UK)	Brain Region Affected	Molecular Changes	Symptoms
Alzheimer's Disease (AD)	225,000	Hippocampus, amygdala, neo- cortex, basal forebrain	 Neurofibrillary tangles (hyper- phosphorylation of tau proteins) β-amyloid plaques (amyloid peptide aggregates) 	Dementia (cognitive, psychiatric, behavioural)
Amyotrophic Lateral Scle- rosis (ALS)	1,300	Motor neurons	· Motor neuron degeneration in spinal cord, brain stem, primary motor cortex	Difficulty walking, weakness in limbs/arms, slurred speech, muscle cramps and twitching
Parkinson's Disease (PD)	60,000	Substantia nigra	 Dopaminergic neuron loss from nigrostriatum and substantia nigra pars Build-up of Lewy bodies, α-synuclein aggregates 	Tremors, bradykinesia, impaired balance and posture, loss of automatic movements
Huntington's Disease (HD)	2,000	Basal ganglia, neostriatum	Trinucleotide (CAG) repeat in ITI5 huntington gene Loss of medium spiny neurons (MSN) Toxicity of mHTT: protein misfolding & aggregation	Muscle problems (rigid- ity, dystonia), involun- tary jerking movements, abnormal eye move- ments
Spinal Mus- cular Atrophy (SMA)	300	Motor neurons	Mutations in SMN1 gene	Muscular atrophy, bal- ance and coordination deficits

Within the context of the two main developing therapies examined, gene and stem cell therapy, I suggest specific focuses for research in their current fields. Major milestones in research that must be reached before clinical application begin with better understandings of disease initiation and progression. Molecular pathological classification through the identification of biomarkers can provide the greatest cross-methodological benefits in furthering effective and personalized treatment of NDs. Fundamentally, a more thorough understanding of underlying ND pathophysiology can be accomplished through biomarker identification, stem cell-based disease modelling, and genetic research. Combined, this knowledge will help develop more personalized and ultimately more effective novel therapies for treating neurodevelopmental disorders.

Part 1: Current Approaches: Disease Management

Section 1.0: Introduction

Current clinical approaches to addressing AD, PD, and HD all center on disease management and symptom alleviation. The use of pharmacological agents, physical therapy, and caregiving aid is efficient and affordable, but are ultimately only forms of palliative care rather than effective treatments.

Section 1.1: Current Management Techniques

Four acetylcholinesterase inhibitors (donepezil, galantamine, rivastigmine, tacrine) and one NMDA receptor antagonist (memantine) constitute the five main medications currently used to provide temporary symptomatic relief to AD patients (Desai & Grossberg, 2005). The first four enhance cholinergic function by reducing rates of acetylcholine breakdown (as reduced ACh levels is related to memory and cognitive decline in AD), while memantine reduces ADrelated excitotoxicity (observed in AD & related to neuronal death). Further behavioural, psychosocial, and caregiving provisions are used to improve quality of life for AD patients.

Current therapies for PD are arguably the most well-developed and effective among these three discussed NDs. Drugs such as levodopa or dopamine agonists used in earlier stages of care can restore some motor function, but their efficacy decreases dramatically over time. Deep brain stimulation (DBS) and rehabilitation techniques provide further symptomatic therapies for PD (Mizuno, 2014).

Treatments for HD, like those for AD and PD, are only able to reduce symptom severity. Various pharmacological treatments are available to reduce Chorea, a disorder within Huntington's characterized

Table 2: Current Therapies for AD, PD, & HD

Disease	Pharmacological Treatments	Other Treatments		
Alzheimer's Disease (AD)	Acetylcholinesterase inhibitors (donepezil, galantamine, rivastigamine, tacrine) NMDA receptor antagonist (memantine)	Caregiving Behavioural therapy (reduce problem behaviours)		
		· Psychosocial interventions (emotion and cognition regulation)		
Parkinson's Disease (PD)	Dopamine replacement (levodopa, carbidopa, selegeline)	Deep brain stimulation (of thalamus, globus pallidus, subthalamic nucleus) Physical therapy		
Huntington's Disease	Chorea treatment (tetrabenazine, benzodiazepines, neuroleptics)	Caregiving Physical therapy Speech and language therapy		

by involuntary spasmodic movements (Busse et al., 2012). As the disease progresses, physical, speech, and language therapies are used to rehabilitate physical and cognitive function (Busse et al., 2012).

Section 1.2: Moving Forward

Techniques currently available to treat neurodegenerative diseases are able to provide temporary symptomatic relief, but are critically unable to halt the rapid deterioration and progression of neurodegeneration.

Though the complete causes of AD and PD have not been fully characterised, neurofibrillary tangles, β-amyloid plagues, decreased synaptic function, and cortical atrophy observed in AD and the loss of dopaminergic neurons characteristic of PD can be addressed by gene therapy and stem cell-based therapies. These two therapies can also be applied to treat the root causes of HD, which has been attributed to an expanded trinucleotide repeats, the mHTT protein, and consequent neostriatal neuronal loss. However, their widespread clinical application has been limited by underdeveloped understandings of disease etiology and mechanisms of therapeutic action. Insights into the molecular mechanisms of NDs lie at the crux for the development of novel targets of disease-modifying therapies. Therefore, efforts to further gene and stem cell-based therapies (discussed in Parts 2 & 3) will benefit from an increased focus on biomarker research (discussed in Part 4) to elucidate disease pathophysiologies.

Part 2: Gene Therapy

Section 2.0: Introduction

The theory behind the use of gene therapy to treat neurodegenerative disease is simple: deliver a transgene that either corrects or replaces a defective gene to cure a complex disease using a transfer vector. However, the numerous technical difficulties associated with gene therapy are daunting in complexity: the transfer vector must be easy to produce, safe for patients and the environment, target specific cells efficiently, and not trigger a deleterious immune reaction; simultaneously, the appropriate transduced transgene must be expressed at the optimal level for

an optimal duration (Sarkis & Mallet, 2005). All these factors that must be taken into consideration can only be optimized with increased knowledge of disease etiology, which can be mediated through biomarker research.

Section 2.1: Gene Therapy Trials

Gene therapy vectors are either viral or non-viral. Currently, the most common viral vectors are adenoassociated viruses (AAVs) and lentiviruses. Both infect non-dividing and dividing cells, but lentiviral vectors integrate into host genomes while AAVs do not. Thus, lentivirus integration can confer long-term, stable transgene expression, but poses a risk of integrational mutagenesis. Non-viral vectors (mostly used for HD treatment) have more localized effects, require higher dosage levels, and confer only transient gene expression that is normally insufficient to fully treat chronic ND.

2.1.1: Viral Techniques

A series of viral vector-mediated gene therapy trials has been conducted for AD and PD, summarized in the table below. In these trials, AAV2-mediated transgenes were injected into patients' brains, but treatment results either fail to meet or achieve the same level of therapeutic benefit as current standards of care. It is important to emphasize that the efficacy of gene therapy depends critically on the therapeutic gene delivered; this can only be improved by better understandings of the etiology of neurodegenerative diseases.

2.1.2: Non-Viral Techniques

Huntington's is a genetically acquired disease with a known etiology (CAG repeats in the ITI5 huntington gene), for which non-viral gene therapy techniques carry significant therapeutic potential. Carroll et al. (2011) have developed antisense oligonucleotides (ASOs) that selectively bind to exonic and intronic SNP (single nucleotide polymorphisms) sites on the mutant huntingtin protein (mHTT), thus selectively silencing gene expression of mutated regions on the mHTT. The experiment used primary cells from HDpatients with transgenic mouse lines, and the devel-

Table 2: Current Therapies for AD, PD, & HD

Disease	Study / Trial Codes	Gene Therapy	Method of Delivery	Current status
Alzheim- er's Dis- ease (AD)	Rafii et al., 2014 (NCT00876863)	AAV2-NGF NGF: nerve growth factor // dys- regulated in AD, neuroprotector/ cellular therapy efficacy	Injection into basal forebrain	· Phase 1 successful*, therapy well-tolerated, NGF-induced axonal sprouting observed · Phase II in progress
Parkin- son's Dis- ease (PD)	LeWitt et al., 2011 (NCT00643890)	AAV2-GAD GAD: glutamic acid decarboxylase // rate-limiting biosynthetic enzyme for GABA, reduces excitoxicity	Injection into sub- thalamic nucleus	Phase I & II successful, but symptom improvements achieved were not better than current standards of care
Parkin- son's Disease	Bartus et al., 2011 (NCT00400634; NCT00985517)	AAV2-NRTN NRTN: neurturin // delivers support to dopaminergic neurons	Injection into striatum & putamen	Phase I successful, observed increased NRTN expression Phase II showed no improvement in clinical outcome over controls
Parkin- son's Disease	Azzouz et al., 2002 (NCT00627588; NCT01856439)	Lentivirus-TH, AADC, GCH1 (*exciting tricistronic vector: cells transduced express all 3 enzymes) TH: tyrosine hydroxylase / AADC: aromatic amino acid dopa decar- boxylase / GCH1: GTP cyclohy- drolase I // three genes encode rate-limiting do- pamine biosynthesis enzymes	Injection into striatum & putamen	Phase I (open label) successful, improved motor function correlating with increasing dose Currently optimizing delivery method before Phase II (double-blinded randomized trial)
Parkin- son's Disease	Muramatsu et al., 2015 (NCT02418598)	AAV2-AADC (as discussed above)	Injection into puta- men	Phase I safety trial successful, stable AADC expression 4 years postdelivery Phase II recruiting, in progress
Parkin- son's Disease	(NCT00229736)	AAV2-AADC	Intrastriatal infusion	· Phase I safety trial suc- cessful

^{*}success defined by meeting safety requirements and showing functional improvement

oped ASOs achieved potent and selective allele-specific knockdown of mHTT both in vitro and in vivo.

This presents the most exciting potential treatment for Huntington's, as ASO delivery to adult CNS can be achieved through simple intrathecal or intracerebroventricular infusion rather than viral delivery. Further, ASO dosing can be precisely controlled or stopped when necessary. Kordasiewicz et al. (2012) have shown that transient infusion of ASOs catalysing RNase H-mediated degradation of mHTT in mouse and non-human primate models delays disease progression and sustains HD disease reversal that outlasts ASO-mediated mHTT knockdown. Thus, clinical trials should be conducted to test the efficacy of gene therapy either through direct ASO infusion, or combined with iPSC-mediated transplantation (correcting disease mutations of patient-derived iP-SCs with ASOs, then transplanting corrected NPCs).

Section 2.2: Conclusion

Promising proof-of-principle concepts have been presented in Phase I trials, which have shown that gene therapy techniques targeting AD & PD are safe and well-tolerated. However, therapeutic efficacy in further clinical trials remains weak, in that observed improvements failed to exceed placebo effect or current standards of care for AD and PD. Non-viral ASOmediated silencing of mHTT expression has proved efficacious in HD-animal models but have not yet moved to clinical trials. In sum, success by design meets failure of efficacy within these gene therapy studies.

Moving forward, future research must focus first on expanding choices of possible transgenes for delivery, as improvements in vectors and delivery methods depend crucially on efficacy of the transgene delivered. Greater understanding in disease etiology will help identify these gene targets. A significant area of development lies in the identification of biomarkers of NDs, which would not only improve gene therapy techniques, but also allow for earlier diagnoses, continuous motoring of the progression of therapeutic effects, and the development of precision medicine.

Part 3: Stem Cell Therapy

Section 3.0: Introduction

Stem cell and iPSC research has significantly furthered neurodegenerative disease modelling, diagnosis, and transplant- and drug-based therapies. A plethora of therapies developed in animal models have failed to translate into effective clinical trials due to differences in mammalian genomes and embryonic development (Begley & Ellis, 2012). Humanbased stem cell research generates not only potential ND therapies, but also presents clearer pathophysiological models for those diseases. In particular, the development of induced pluripotent stem (iPS) cells has presented a unique approach for studying signalling pathways, growth control, and disease mechanisms in previously inaccessible human brain tissue.

The main types of stem cells used for neurodegenerative therapies are embryonic, mesenchymal, progenitor, and iPS cells (Singh et al., 2016). Embryonic stem cells are pluripotent and hold excellent potential to restore damage caused by brain injury or neurodegeneration; however, their ability for unlimited self-renewal poses a high risk for tumour formation post-engraftment, limiting the range of their clinical application. Mesenchymal stem cells are multipotent and immunomodulating (derived from autologous source; do not trigger a host immune response), and thus find a relatively widespread application for neurodegenerative treatment. However, their use in genetic diseases, wherein autologous sources contain the same genetic predisposition to the disease, is limited. Neural progenitor cells (NPC) and neural stem cells (NSC) are derived from either fetal or adult neural tissue, carry limited differentiation and tumour-formation potentials, but are difficult to isolate and thus lower in availability. iPSCs, generated from somatic tissue and reprogrammed using TFs into embryonic stem cell-like states, offer novel sources of autologous cellular therapy.

Application of the appropriately selected type of stem cell to treat neurodegenerative diseases is mediated through two primary mechanisms. First, stem cells can replace or stabilize neuronal networks that experience both widespread or specific subpopulation-based neuronal loss. Second, stem cells can provide environmental enrichment through neurotrophic support to prevent further neuronal degradation. These mechanisms will be discussed in the context of specific neurodegenerative diseases below. Following this discussion, I'll emphasize that the focus of stem cell research should first lie in exploiting its means of modelling disease pathophysiology and providing an in vitro screening tool for screening efficacy and safety of drugs, then on its direct therapeutic applications. I'll then highlight the potential iPSC research holds in future developments of precision medicine.

Section 3.1: Alzheimer's

Complementing current pharmacological AD treatments that regulate neurotransmitter activity (Section 1.1), stem cell (SC)-based therapies for AD can also help restore degeneration and loss of cholinergic neurons, and increase levels of brain-derived neurotrophic factor (BDNF) and nerve growth factor (NGF) that are lowered in AD. In mouse models, combinatorial upregulation of BDNF with NPC transplants promoted neurogenesis and protected neuronal function. Thus, NPC-mediated stem cell-based therapy may offer an enhanced approach to preventing AD-associated degradation of cholinergic synapses and neurons (Tuszynski, 2007).

Wang et al. (2006) compared transplants of ESC-derived NSC to transplants of ESC alone into mouse AD models. As suggested in Section 3.0, ESC-transplanted mice developed teratomas (ESCs: unlimited selfrenewal capabilities, high risk of tumour formation), while transplanted NSCs successfully differentiated into stable cholinergic neurons, improving memory and learning. Genetically modified NSCs for BDNF expression has been shown in many rodent studies to reduce AD symptoms (Lindvall & Kokaia, 2010). In these mouse models, increased hippocampal synaptic density, increased cognitive function, and enhanced cell-cell synaptic communication, as associated with increased BDNF expression (Blurton-Jones et al., 2009; Li et al., 2009; Xuan et al., 2008). NGF has also been implicated in preventing neurodegeneration and reducing amyloid toxicity (Tuszynski et al., 2007). In addition to AAV2-mediated gene therapy techniques discussed in Section 2.1.1, autologous fibroblasts genetically modified to express NGF implanted into patient forebrains have been found to slow rates of cognitive decline in an open-label phase 1 clinical trial (Tuszynski et al., 2005). However, fibroblasts are immobile and cannot slow degeneration

across large brain regions. Transplanted stem cells can efficiently migrate and release growth factors to many damaged sites, but their use in AD treatment has yet to be fully explored. Another technique to increase NGF release to treat AD is with encapsulated cell biodelivery (ECB) implants, for which the first clinical trial was conducted in 2008 by the Danish company NsGene. Encapsulated retinal pigment epithelial cells releasing NGF were implanted into the basal forebrains of six AD patients (Wahlberg et al., 2012). At 12 months, implants were retrieved and low but persistent NGF secretion was detected in half of the subjects. Thus, both ECB implantation and stem cell-based gene therapy may provide future methods for enhancing NGF expression to counteract cholinergic neuronal death.

Although NSC transplantations into the adult forebrain of patients may be able to increase cholinergic and cognitive aptitude, they are unable to target the main molecular cause of the disease: accumulation of β-amyloid plagues (Blurton-Jones et al., 2009). Nonetheless, the ability of NSC transplants to enhance cholinergic function, synaptic transmission, and neuronal function is suggested by animal studies and replicated by initial clinical trials (Tuszynski, 2007). Combining growth factor overexpression with NPCs' ability to integrate across many neural networks thus presents an exciting potential for limiting AD pathogenesis. Continued research must be conducted before established SC-based treatments can be developed for AD therapy.

Section 3.2: Parkinson's

Earlier open-label studies of human fetal ventral mesencephalic tissue (hfVM) transplants in Parkinson's patients yielded successful long-term PD symptomatic relief, but these results failed to replicate in subsequent placebo-controlled, double-blind trials (Brundin et al., 2010). Success has been observed in using genetically modified ESCs and iPSCs to derive neurons with the dopaminergic (DA) phenotype, but these techniques have not yet been translated into clinical studies.

Clinical trials using hfVM tissue transplants in the 1990s (Madrazo et al., 1988; Lindvall et al., 1990) found impressive restoration of dopaminergic neurotransmission and motor-symptom alleviation. In these open-label clinical studies, PET scans showed long-lasting evidence of functionally-integrated dopaminergic neurons forming from transplanted tissue, restored DA release in vivo, and reactivation of cortical motor areas. However, the design of these open-label trials contains significant bias and thus deserve cautious interpretation. Further, the occurrence of recurring graft-induced dyskinesias (GIDs) in consequent double-blind trials, in addition to heavy ethical concerns raised over the use of fetal tissue as a therapeutic source, has heavily impeded the clinical development of this form of hfVM-based therapy (Brundin et al., 2010). Instead, the focus has been shifted to ESCs and iPSCs: mouse model studies have found that overexpression of Nurr1, FGF-8, and Shh signalling in mouse ESCs and iPSCs has successfully induced midbrain dopaminergic phenotypes and improved functional recovery (Kim et al., 2003; Cooper et al., 2010). A major concern for ESC clinical transplants, however, is the risk for tumour formation observed in animal models (Lindvall & Kokaia, 2010). As PD patients have normal life expectancies, even a slim risk of tumour formation is unacceptable.

Initial suggestions of hfVM transplant-mediated PD therapy have been significantly hindered by ethical and methodological issues, while the induction of ESCs and iPSCs into dopaminergic neurons is cumbersome, potentially nonhomogeneous (may contain traces of undifferentiated cells that can lead to teratomas), and currently unsubstantiated by clinical studies. Thus, progress made in stem cell-based PD treatments have been slower than for AD. However, iPSC-derived dopaminergic neurons have provided critical insights into the pathophysiology of PD, by providing a SC-based human disease model. Abnormalities in mitochondrial and dopamine homeostasis, as well as mutations in Parkin and PTEN-induced putative kinase 1 (PINK1) previously noted in animal models have been substantiated by similar data from studies using PD patient-derived iPSC neurons (Van Laar et al., 2010; Cai et al., 2012).

Though stem cell technology has yet to develop clinically-relevant PD therapies, it has proved effective in modelling Parkinson's. In particular, mutationbearing iPSCs has helped study PD physiology, elucidate underlying signalling cascades modulating the disease, identify PD biochemical markers, and screen

efficiency of therapeutic compounds.

Section 3.3: Huntington's

Applications of stem cell techniques in HD studies have paralleled those for AD and PD. More specifically, fetal striatal grafting into HD patients have provided transient symptom improvements, but its mechanisms are poorly understood and its use is limited by ethical constraints (Bachoud-Lévi et al., 2000). Both iPSCs and ESCs have been used to establish differentiation protocols for medium spiny neurons (MSN)— these transplantations have shown successful neural circuit development and integration, but are currently restricted to animal models (Viegas et al., 2011; Precious & Rosser, 2012).

Initial clinical trials using fetal-derived tissue, iPSCs, or ESCs as sources of MSNs have observed reduced motor and cognitive dysfunction in HD patients. In addition, decreased neuronal loss was observed in HD rodents with NPC transplants engineered to overexpress GDNF, as compared to control mice with unmodified NSC transplants, emphasizing the potential of stem cell-based therapies to provide environmental enrichment and protect endogenous neurons (Pineda, 2007).

Section 3.4: Conclusion

For AD, PD, and HD, similar trends in stem cellbased therapies can be observed. Early cell-based therapies using fetal ventral mesencephalic tissue (for PD) and fetal striatal tissue (for HD) transplants provided initial support for successful symptomatic relief. However, widespread clinical applications of human fetal tissue transplants has been limited by significant ethical and scarcity issues associated with the nature of the tissue source. Subsequent clinical trials have failed to consistently replicate initial data and observed deleterious side effects (teratoma formation, GIDs, immunorejection). Thus, the focus of stem cell therapies has shifted to ESCs derived NSCs induced to certain cell fates or gene expressions (increased BDNF/NGF expression for AD, dopaminergic phenotypes for PD, and MSN for HD); however, these induced populations are usually not pure, long-term graft survival is generally low, and any ES cells escaping in vitro differentiation carries the potential or yielding teratomas and tumours following transplan-

Table 4: Overview of Current Literature on SC-based ND therapies

Disease	Stem Cell Applications	Goal of SC-based Therapies	Current Therapeutic Progress	
Alzheimer's disease	· NSCs · ESCs	· Restore loss of cho- linergic neurons	· Successful basal forebrain grafts of NGF-secreting fibroblasts (open-label trial, Tuszynski, 2007)	
		· Increase BDNF & NGF trophic factors to pro- mote neurogenesis	· Development of encapsulated cell biodelivery implants to increase NGF release (NsGene)	
			· No current clinical trials testing SC-transplants in AD	
disease		Restore nigrostriatal dopaminergic neuro- nal loss	· Successful clinical open-label hfVM transplants (Lindvall et al., 1990; Madrazo et al., 1988) — pro- vided long-term symptomatic relief, but failed to replicate in subsequent double-blind trials (which observed GIDs)	
			· Successful animal studies using ESCs & iPSCs to derive DA phenotype, not yet translated to clinical trials due to risk of tumour formation and technical difficulties	
Huntington's disease	· MSNs	· Restore medium spiny neuron loss	· Successful clinical fetal striatal grafting (Bachoud- Lévi et al., 2000)— provided transient symptom improvements, but limited by ethical constraints & poorly-understood mechanism of action	
			· Successful animal studies using ESCs & iPSCs to restore MSN loss, not yet translated to clinical trials	

tation.

Above all, the development of effective stem cellbased therapies is ultimately hindered by our current lack of knowledge on the mechanisms of action of transplanted stem cells, and how stem cell proliferation, differentiation, survival, and migration are controlled in a pathological environment. The mechanisms of cellular protection provided by SC-mediated trophic support are also not fully understood (McBride et al., 2004). Because current potential risks outweigh validated benefits in their therapeutic applications, research on human stem cells and patient-derived iPSCs should first be focused on disease modelling and used for screening drugs and other therapeutics. Successful development of clinical therapies can begin once more precise disease pathophysiologies and mechanisms of SC action post-transplantation are identified.

In sum, the use of stem cells for transplantation has generated enormous ethical, technical, efficacy, and safety-related debates. It is true that SC replacement therapy can stabilize and regenerate entire networks

of neuronal loss and prevent further neurodegeneration by providing environmental and trophic factors against toxic, neurodegenerative factors. However, with current literature, the greatest functionally validated potential of SCs lies in their unique ability to model disease pathophysiology and mechanisms of action. Patient-derived iPSC cells may be a particularly tremendous treatment option for personalized medicine in the future, but for now serve as prominent tools for understanding disease etiology on a highly personalized level, as they carry the patient's genotype, disease mutations, and also account for environmental influences on the individual's disease.

Part 4: Precision Medicine & Biomarkers

In discussing future developments of neurodegenerative therapies, it is critical to emphasize the importance of elucidating the molecular etiologies of NDs. Pathologically altered proteins and protein aggregations are fundamental characteristics of neurodegenerative diseases and should be of paramount focus for further research.

I've alluded to the importance of biomarker research in aiding developments of novel therapies during Part 2 & 3's discussion of gene therapy and stem-cell based treatments. This is because molecular classifications of NDs can detail the nature of pathologically dysfunctional proteins and correlate corresponding clinical symptoms to genetic abnormalities. Ongoing biomarker research has already identified extracellular deposits of A fibrils and aggregation of abnormally-phosphorylated tau proteins in AD, the presence of Lewy body deposits, altered dopamine transporters in PD, and protein misfolding and aggregation associated with mHTT proteins in HD (Kovacs, 2016). Thus, the ability of biomarker data to identify core disease pathology has already been validated, and therapies targeting these protein abnormalities (through immunotherapy, gene therapy, SC-transplants) have yielded promising results in animal and some clinical models. However, the most exciting potential of biomarker research lies in its ability to characterise patient-specific environment-disease dynamics and gene variations, and consequently provide personalized diagnoses. In the emerging era of precision medicine, the use of biomarkers (detected in body fluids or PET imaging) can provide earlier and more accurate ND diagnosis, and their identified protein dysfunctions can be targeted in translational research using novel therapies.

Looking forward, biomarker research will contribute greatly to developing pathophysiological disease models, improving immunotherapy, gene therapy and SC-based transplant methods, and moving us closer to personalized cures for neurodegenerative diseases. Similarly, stem cell research can be applied to experimentally model NDs. Patient-derived iPSCs are especially well-suited to uncover specific disease pathophysiologies as they carry the patient's genotype and bear patient-specific disease mutations and gene-environment interactions. Thus, I believe that research focusing on biomarkers, patient-derived iP-SCs, and genetic studies are currently most necessary and potentially most beneficial for current experimental and clinical endeavours for ND cures.

References

Azzouz M., Martin-Rendon E., Barber R.D., Mitrophanous K.A., Carter E.E., Rohll J.B., Kingsman S.M. (2002). Multicistronic lentiviral vectormediated striatal gene transfer of aromatic L-amino acid decarboxylase, tyrosine hydroxylase, and GTP cyclohydrolase I induces sustained transgene expression, dopamine production, and functional improvement in a rat model of Parkinson's disease. J. Neurosci., 22 (23), 10302-10312.

Bachoud-Lévi A-C, Rémy P, Nguyen J-P, Brugières P, Lefaucheur J-P, Bourdet C, Baudic S, Gaura V et al (2000) Motor and cognitive improvements in patients with Huntington's disease after neural transplantation. Lancet 356(9246):1975-1979

Bartus, R.T., Brown L., Wilson, A., Kruegel, B., Siffert, J. Properly scaled and targeted AAV2-NRTN (neurturin) to the substantia nigra is safe, effective and causes no weight loss: support for nigral targeting in Parkinson's disease. Neurobiol. Dis., 44 (2011), 38-52.

Blurton-Jones, M., Kitazawa, M., Martinez-Coria, H., Castello, N. A., Müller, F.- J., Loring, J. F., et al. (2009). Neural stem cells improve cognition via BDNF in a transgenic model of Alzheimer disease. Proc. Natl. Acad. Sci. U.S.A. 106, 13594-13599.

Brundin, P., Barker, R., Parmar, M. (2010). Neural grafting in Parkinson's disease: problems and possibilities. Progress in Brain Research, Vol. 184, Chapter 14.

Busse, M., Khalil, H., Brooks, S., Quinn, L., Rosser, A. (2012). Practice, Progress and Future Directions for Physical Therapies in Huntington's Disease. Journal of Huntington's Disease. 1(2), 175-85.

Carroll, J. B., Warby, S. C., Southwell, A. L., Doty, C. N., Greenlee, S., Skotte, N., et al. (2011). Potent and selective antisense oligonucleotides targeting single-nucleotide polymorphisms in the Huntington disease gene/allele specific silencing of mutant huntingtin. Mol. Ther. 19, 2178-2185.

Chmielnicki, E. & Goldman, S. Stem cells and cell-based therapy in neurodegenerative disease. In Beal, M., Lang, A., Ludolph, A. (Eds.), Neurodegenerative Diseases: Neurobiology, Pathogenesis and Therapeutics (329-40). Cambridge University Press.

Cooper O, Hargus G, Deleidi M, et al. (2010). Differentiation of human ES and Parkinson's disease iPS cells into ventral midbrain dopaminergic neurons requires a high activity form of SHH, FGF8a and specific regionalization by retinoic acid. Mol Cell Neurosci. 45:258–266.

Desai, A. & Gossberg, G. (2005). Diagnosis and treatment of Alzheimer's disease. Neurology. 64(12), S34-9.

Ebert AD, Barber AE, Heins BM, Svendsen CN. Ex vivo delivery of GDNF maintains motor function and prevents neuronal loss in a transgenic mouse model of Huntington's disease. Experimental neurology. 2010; 224:155-162.

Goldman, S. (2005). Stem and progenitor cell-based therapy of the human central nervous system. Nature Biotechnology. 23(7), 862-70.

Li G, Peskind ER, Millard SP, et al. (2009). Cerebrospinal fluid concentration of brain-derived neurotrophic factor and cognitive function in non-demented subjects. PLoS One. 4:e5424.

Lindvall O, Brundin P, Widner H, Rehncrona S, Gustavii B, Frackowiak R, Leenders KL, Sawle G et al. (1990). Grafts of fetal dopamine neurons survive and improve motor function in Parkinson's disease. Science. 247(4942):574-577

Lindvall, O., Kokaia, Z. (2010). Stem cells in human neurodegenerative disorders—time for clinical translation? Journal of Clinical Investigation. 120(1), 29-31.

Kim T. E., Lee H. S., Lee Y. B., Hong S. H., Lee Y. S., Ichinose H., et al. . (2003). Sonic hedgehog and FGF8 collaborate to induce dopaminergic phenotypes in the Nurr1-overexpressing neural stem cell. Biochem. Biophys. Res. Commun. 305, 1040-1048. 10.1016/S0006-291X(03)00879-9

Kordasiewicz, H. B., Stanek, L. M., Wancewicz, E. V., Mazur, C., Mcalonis, M. M., Pytel, K. A., et al. (2012). Sustained therapeutic reversal of Huntington's disease by transient repression of huntingtin synthesis. Neuron 74, 1031–1044.

Kovacs, G. (2016). Molecular Pathological Classification of Neurodegenerative Diseases: Turning towards Precision Medicine. Int. J. Mol. Sci. 17(189), 1-21.

Kumar, A., Narayanan, K., Chaudhary, R., Mishra, S. 2016. Current Perspective of Stem Cell Therapy in Neurodegenerative and Metabolic Diseases. Mol. Neurobiol., online.

Sarkis, C. & Mallet, J. (2005). Gene Therapy. In Beal, M., Lang, A., Ludolph, A. (Eds.), Neurodegenerative Diseases: Neurobiology, Pathogenesis and Therapeutics (329-40). Cambridge University Press.

Singh, S., Srivastava, A., Srivastava, P., Dhuriya, Y. 2016. Advances in Stem Cell Research- A Ray of Hope in Better Diagnosis and Prognosis in Neurodegenerative Diseases. Frontiers in Molecular Biosciences. 3(72), 1-10.

Tuszynski, M. H. (2007). Nerve growth factor gene therapy in Alzheimer disease. Alzheimer Dis. Assoc. Disord. 21, 179–189.

Tuszynski, M. H., Thal, L., Pay, M., Salmon, D. P., Bakay, R., Patel, P., et al. (2005). A phase 1 clinical trial of nerve growth factor gene therapy for Alzheimer disease. Nat. Med. 11, 551–555.

Mizuno, Y. (2014). Recent Research Progress in and Future Perspective on Treatment of Parkinson's Disease. Integr Med Int. 1(2), 67-79.

Rafii, M.S. et al. (2014). A phase1 study of stereotactic gene delivery of AAV2-NGF for Alzheimer's disease. Alzheimers Dement., 10, pp. 571–581

McBride JL, Behrstock SP, Chen EY, et al. Human neural stem cell transplants improve motor function in a rat model of Huntington's disease. J Comp Neurol. 2004; 475:211–219.

O'Connor, D. & Boulis, N. (2015). Gene therapy for neurodegenerative diseases. Trends in Molecular Medicine. 21(8), 504-11.

Precious SV, Rosser AE (2012) Producing striatal phenotypes for transplantation in Huntington's disease. Exp Biol Med 237(4):343–351

LeWitt, P. et al. (2011). AAV2-GAD gene therapy for advanced Parkinson's disease: a double-blind, sham-surgery controlled, randomised trial. Lancet Neurol., 10, pp. 309–319

Xuan AG, Long DH, Gu HG, et al. (2008). BDNF improves the effects of neural stem cells on the rat model of Alzheimer's disease with unilateral lesion of fimbria-fornix. Neurosci Lett. 440:331–335.

Viegas P, Nicoleau C, Perrier AL (2011) Derivation of striatal neurons from human stem cells. Prog Brain Res 200:373–404

Wahlberg, L. U., Lind, G., Almqvist, P. M., Kusk, P., Tornøe, J., Juliusson, B., ... Linderoth, B. (2012). Targeted delivery of nerve growth factor via encapsulated cell biodelivery in Alzheimer disease: A technology platform for restorative neurosurgery. Journal of Neurosurgery, 117(2), 340-347. doi:10.3171/2012.2.jns11714